Comparison of carbon stores by two morphologically different seagrasses *

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INTRODUCTION

The recent emphasis on global change has intensified research in the carbon sequestration potential of seagrass ecosystems (Nellemann et al. 2009). Most estimates on seagrass carbon storage are, however, derived from studies of a few species and habitats. Lavery et al. (2013) showed an 18-fold difference in carbon stores among Australian seagrasses highlighting the importance of inter-habitat variability in carbon stocks. One implication from this is that the factor of seagrass species may play a key role in the variation in carbon stores beneath those studied meadows. Different seagrass species have inherently dissimilar traits, well summarised by the seagrass functional-form model in Carruthers et al. (2007). This functional-form model differentiates seagrasses based on morphological plasticity, rhizome persistence and occurrences in varying depositional environments, among other traits. A further extension of this model is suggested such that the seagrass species may shift from low to high transitions of standing crop biomass between extreme ends of the model. Of interest, the two seagrass genera Halophila and Posidonia are placed at opposite ends of the functional-form model. Halophila has a small biomass with less persistent rhizomes while Posidonia has the opposite traits. We hypothesise that meadows of smaller, ephemeral seagrasses with low standing crop will have less accumulated carbon compared to the larger and more persisting forms with higher productivity and biomass. Estuarine seagrass habitats dominated by Halophila ovalis and Posidonia australis were studied to compare the total carbon stocks and origin of the preserved carbon.

METHODS

Four seagrass meadows were studied: *P. australis* from Oyster Harbour (OH: 34°58'58.3"S, 117°58'29.9"E) and Waychinicup Inlet Estuary (WI: 34°53'35.9"S, 118°19'57.7"E), and *H. ovalis* from Swan River Estuary (SR: 32°00'46.1"S, 115°47'43.5"E) and Harvey Estuary (HE: 32°38'00.3"S, 115°38'51.7"E). To allow viable comparisons

not confounded by habitat types potentially influencing the depositional environment of allochthonous carbon, this study compared estuarine meadows of both species. Both tidal and swell exchanges dominate WI while OH, SR and HE are wave dominated. Three sediment cores were each collected from the four sites in 2012 by either manually hammering core barrels or by vibracoring (Vibecore-D, SDI) into the sediments. These cores were sliced, dried and weighed for dry bulk-density analysis. Alternate slices of ground and acidified (1 M HCl) subsamples were encapsulated and analysed for total organic carbon (OC) and stable carbon isotope composition values (813C) at the UC Davis Stable Isotope Facility (continuous flow isotope ratio mass spectrometer analyser, Sercon). For testing differences in sedimentary OC content among different seagrass species and sites, a two-way nested ANOVA (SPSS 19) was applied (species as fixed factor, site nested within species) and post-hoc (Tukey) tests were further applied to assess differences among sites within the same species.

RESULTS

The sampling resulted in different lengths for all cores. After corrections for sediment compression during coring (Glew et al. 2001), core lengths ranged from 170 cm to 250 cm. To allow comparisons, OC characteristics in all sediment cores were standardised for the top 170 cmthick deposits. In the Posidonia cores, localised agglutination of plant detrital matter was observed. This coarse Posidonia detritus was more abundant in OH cores. The top 10 cm of Halophila cores contained low amounts of coarse plant detrital matter becoming less evident until the 25 cm depth and was absent below this level. Mean OC content was higher in P. australis sites (mean±SE; 2.03±0.19% at OH and 1.12±0.08% at WI) compared to H. ovalis sites (0.34±0.15% at HE and 0.16±0.03% at SR). The mean OC content in 170 cm-thick deposits was 6-fold higher in P. australis sites (1.58±0.21%) compared to H. ovalis sites (0.25 \pm 0.07%; p<0.05). The OC content was significantly higher in P. australis OH cores compared to those in W1 (p<0.01), while the OC content among H. ovalis sites was similar (p>0.05). δ^{13} C values of sedimentary organic matter in Posidonia meadows at OH were more positive (-9.89‰ to -14.0‰) than in WI (-13.0% to -19.5%). The δ^{13} C signatures in both *Halophila* sites followed similar trends in all cores (ranging from -16.0% to -21.5%).

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DISCUSSION

Comparison of carbon stocks between species

The OC stocks in 170 cm-thick deposits beneath P. australis meadows were 6-fold higher than in H. ovalis sites. The difference in OC accumulation between the two different seagrass species can be partially explained by the higher biomass and productivity of P. australis meadows (1300-2100 g DW m⁻² and 0.23 g DW m⁻² d⁻¹, respectively) compared to the H. ovalis meadows (76 g DW m⁻² and 0.01 g DW m⁻² d⁻¹: Duarte & Chiscano 1999). In addition, Posidonia species invests larger amounts of carbon into below-ground organs (1220 g DW m⁻²) compared to small seagrasses such as Halophila (21 g DW m-2), which invests more energy in rapid clonal propagation resulting in high turnover rates. The belowground organs of seagrasses are more recalcitrant than above-ground organs, and therefore are more likely to end buried in situ as detrital matter, particularly for P. australis rhizomes which can grow 20-50 cm beneath the sediment surface. Only in the event of major scouring would these below-ground tissues be uprooted and exported. Furthermore, the dense and relatively high canopy of P. australis stabilises the sediment, enhancing sediment deposition and reducing re-suspension, and increasing the likelihood of OC stocks being preserved. This contrasts with H. ovalis meadows with a sparse and low canopy. Below-ground living organs are ephemeral growing at 5-10 cm below the sediment surface. Any detrital matter produced by H. ovalis is thus more likely to be scoured and allochthonously transported rather than remain buried in situ.

Source of carbon

The δ^{13} C values of sedimentary organic matter in P. australis sites (mean = -13.82% for both sites) are indicative of a strong influence of P. australis-derived input. The reported δ^{13} C values of *P. australis* organs range from -9.9% to -11.9% (Hemminga & Mateo 1996), consistent with those observed in the deposits. In contrast, δ^{13} C values at *H. ovalis* sites (mean = -19.16%) are indicative of low amounts of seagrass detritus contributing to the sedimentary organic pool. The reported δ¹³C values of *H. ovalis* organs range from -6.4‰ to -15.5% (Hemminga & Mateo 1996), much more enriched than those we observed in the sediments. The δ¹³C values of allochthonous sources of organic matter (i.e. seston, algae and terrestrial organic matter) are more negative than seagrass isotopic signatures, in the range of -13‰ to -29% (Smit et al. 2005; Loneragan et al. 1997) and may account for the relatively depleted values in the H. ovalis sediments. Previous studies have commented on the ability of the Halophila canopy to effectively trap allochthonous carbon (Fonseca 1989) and our findings are consistent with an earlier study which found that H. ovalis had the second highest OC accumulation after P. australis meadows, possibly due to the nature of the depositional environment and canopy-trapping of allochthonous carbon (Lavery et al. 2013).

Within-species variation

Both *H. ovalis* sites showed similar vertical profiles in the mass and δ^{13} C values of buried OC. In contrast, while we expected the two *P. australis* sites to show similar vertical

OC content trends and $\delta^{13}C$ values through the cores, OH meadows contained double OC stocks of mainly seagrass-derived OC compared to WI. Subsequent radiocarbon dating of the cores from WI (unpubl. data; accelerator mass spectrometry, Australian Nuclear Science and Technology Organisation) showed that at similar stratigraphic levels, the organic matter age varied by ~1000 calibrated years before present. A plausible explanation for this variation in the chronostratigraphy among cores is a scouring event leading to major sediment reworking at WI resulting in a loss of buried OC through erosion. Sediment scouring may also occur in estuaries with higher degree of tidal and swell influence, or during extreme events, and thus could affect the sedimentary OC and δ^{13} C characteristics.

CONCLUSIONS

The comparison of OC stocks and δ¹³C signatures of sedimentary organic matter in the two morphologically different seagrass species demonstrated significant variations in both the carbon accumulation potential as well as the origin of buried carbon among the seagrasses. Meadows of the larger P. australis accrue 6-fold more OC than those of the smaller H. ovalis species. Seagrassderived organic matter forms the bulk of those higher stores while allochthonous organic matter is the major contributor in the Halophila sites. However, within sites of the same species, further variations in carbon characteristics may be exhibited due to natural ecological processes. The results of this study clearly confirm that both species and habitat may contribute to variation in the OC stored in seagrass meadows and that more comprehensive scrutiny of the factors accounting for those variations are required to improve global estimates of Blue Carbon storage in seagrass meadows.

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REFERENCES

CARRUTHERS T J B, DENNISON W C, KENDRICK G A, WAYCOTT M, WALKER D I & CAMBRIDGE M L 2007. Seagrasses of south-west Australia: a conceptual synthesis of the world's most diverse and extensive seagrass meadows. *Journal of Experimental Marine Biology and Ecology* 350, 21–45.

DUARTE C M & CHISCANO C L 1999. Seagrass biomass and production: a reassessment. *Aquatic Botany* **65**, 159–174.

Fonseca M S 1989. Sediment stabilization by Halophila decipiens in comparison to other seagrasses. Estuarine, Coastal and Shelf Science 29, 501–507.

GLEW J, SMOL J & LAST W 2001. Sediment core collection and extrusion. In: Last W M & Smol J P (eds) Tracking

- environmental change using lake sediments, pp. 73–105. Kluwer Academic Publishers, Dordrecht.
- LAVERY P S, MATEO M-Á, SERRANO O & ROZAIMI M 2013. Variability in the carbon storage of seagrass habitats and its implications for global estimates of Blue Carbon ecosystem service. *PLoS ONE* 8(9): e73748. doi:10.1371/journal.pone.0073748.
- LONERAGAN N R, BUNN S E & KELLAWAY D M 1997. Are mangroves and seagrasses sources of organic carbon for penaeid prawns in a tropical Australian estuary? A multiple stable-isotope study. *Marine Biology* **130**, 289–300.
- NELLEMANN C, CORCORAN E, DUARTE C M, VALDÉS L, DE YOUNG C, FONSECA L & GRIMSDITCH G 2009. Blue Carbon. A Rapid Response Assessment. United Nations Environment Programme, GRID-Arendal.
- SMIT A J, Brearley A, Hyndes G A, Lavery P S & Walker D I 2006. Carbon and nitrogen stable isotope analysis of an *Amphibolis griffithii* seagrass bed. *Estuarine Coastal and Shelf Science* 65, 545–556.